

ISSN: 0973-4929, Vol. 17, No. (3) 2022, Pg. 765-794

Current World Environment

www.cwejournal.org

Status and Scope of Conventional Morphometry and its Integration with Bar Coding in Jammu and Kashmir (J&K) Fisheries.

ANCHAL CHIB and SHVETAMBRI JASROTIA*

Department of Zoology, Central university of Jammu, Rahya Suchani, Samba.

Abstract

Correct identification is at the core of taxonomy. Although morphometric characterization is the most straightforward and cost-effective method, it has a number of limitations. By overcoming this limitation, bar coding of mitochondrial cytochrome c oxidase gene (COI) helps in accurate and cost-effective identification of fish species. This communication discusses limitations of conventional morphometry and how its integration with bar coding can help to solve the taxonomic ambiguity of morphologically similar species. Along with that information, different water bodies from J&K region have been analysed with special reference to fish diversity which revealed that the diversity is more in Jammu region as compared to Kashmir, due to favourable climatic conditions and larger number of lotic water bodies and also the fact that the integrated approach has been successfully adopted in Kashmir valley, contributing to accurate identification of fish fauna. There is a lot of room for research in this area as bar coding of fishes has just become popular in the Jammu region, despite its immaturity. This will aid in a better knowledge of the region's fish ecology, the preservation of the gene pool, and the rise of economically significant species.



Article History

Received: 01 June 2022 Accepted: 21 September 2022

Keywords

Anthropogenic Stress; Coi: Gene Pool Conservation; Icthyo Faunal Diversity; Water Bodies of J&K.

Introduction

The ichthyofauna found in riverine ecosystems provides essential supplies. Therefore, an important strategy for future sustainable use and conservation management of both the species and aquatic ecosystems is knowing the Ichthyofaunal population structure.1

Because it serves as the basis for all other life sciences, taxonomic clarity is a vital prerequisite. As sampling and identification are the initial stages, therefore it is the duty of a researcher to precisely identify a species for the purpose of conservation and sustainable use. In recent times, biodiversity research is under priority and new fish identification

CONTACT Shvetambri Jasrotia 🔀 chibaanchal@gmail.com 🛇 Department of Zoology, Central university of Jammu,Rahya Suchani,Samba.



© 2022 The Author(s). Published by Enviro Research Publishers.

This is an f Open Access article licensed under a Creative Commons license: Attribution 4.0 International (CC-BY). Doi: http://dx.doi.org/10.12944/CWE.17.3.23

techniques have been developed, however, their practical application during fish identification is still in its infancy. The most popular, straightforward, economical, and historical approach of identifying fish habitats is morphometric characterization.^{2,3,4}

Morphometric Characterization

All life forms express their phenotypic characters as a result of their genetic constituents and various environmental influences. Morphometrics is basically a more or less interwoven set of statistical procedures used to analyze variability in size and shape measurements of organs and organisms. Morphometric characterization is an old traditional practice being used in fisheries science since 1980's, therefore there is huge array of literature available to enhance pertinent knowledge regarding this.⁵

In order to identify a species of fish, morphological characteristics such as body shape, scale size and count, colour pattern, number and relative location of fins, type and number of fin rays, and numerous relative measures of body parts are analysed.⁶ The statistical analysis of various morphometric characters has helped in recognition of various fish samples.⁷

The morphometric characterization involves the analysis of both measureable and countable characters i.e. meristic characters.⁸ Even though the meristic characters provide some evidence for stock separation, morphometric characters provide the best statistical separation.⁹ Analysis of morphometric characters help not only in correct identification of fishes but also in analysis of migration pattern, stock discrimination, reproductive isolation and commercially important species.¹⁰

Although external morphological characteristics are usually used to identify fish species, there are many distinct fish species and their many developmental phases that make it challenging to do so.¹¹

Fish Habitat and its Ecology Plays A Major Role in Influencing its Morphometric Characterization. A. Resource Specialization and Ecological Conditions of Habitat

Fish exhibit the phenomenon of morphological adaptation depending on the resources and ecological conditions of their habitat, the availability of food, the temperature etc. As a result, it is possible for the same fish raised under different ecological conditions to develop different phenotypic characteristics.^{12,13,14,15,16,17} hence there is a risk of misidentification if visual assessment is employed to identify fishes.¹⁸ Convergent and divergent adaptations also affect the correct identification.¹⁹

B. Morphologically Identical Species

In certain cases using morphometry for species identification yields errornous results because of close resemblance between the morphometric characters,²⁰ sometimes lack of quality in original description can also lead to errornous results.²¹

C. Early Life Stage

Morphological identification of eggs and larval fishes is more difficult, as their morphometric characters are not fully developed.^{22,23} Morphometric characters are also subject to ontogenic transformation leading to error as they change during the process of development.²⁴

D. Sex of the Fish

Although it has been seen that the sex does not significantly influence the morphometric or meristic characters²⁵ but error in morphological identification also depends on the sex of the fish eg. female sharks are more prone to misidentification as compared to male sharks, also the error rate is inversely propotional to body length.²⁶

E. Mislabelled Fishes

Fishes identified by conventional methods being sold in market couldbe mislabeled, either intentionally to fetch higher prices or unintentionaly due to close resemblance between species, incorrect identification of edible fishes can lead to fluctuations in market prices, also some times the mislabeled fishes being sold in the market could be poisonous.^{27,28,29} Therefore correct identification of fishes is essential to prevent their mislabeling.³⁰ Also traditional methodology helps inidentifying the live or dead fish in good form, but not applicabe for identification of processed or mixed samples.³¹

F. More Intraspecific Variations than Interspecific Variations

Even with whole fish specimens, morphometric characterization is occasionally not a good enough

option because they can display either more intraspecific variations or minor interspecific variations. For example, it can be challenging to distinguish between the various Barbus species that live in the Iberian Peninsula based solely on external morphology.³²

G. lack of Classical Taxonomists and Pertinent Literature

Taxonomists provide crucial knowledge about ecosystem thereby providing the key information in life sciences. It has been estimated that about 6000-10000 taxonomists are working worldwide with only a few of them are from developing countries that inhabit most of the Earth's biodiversity.^{33,34} This limited taxonomic community's distribution of competence is similarly uneven; more than 80% of taxonomists are either close to or older than 50 years of age, many among which are not having much computer knowledge.³⁵ Therefore not able to send or retrieve literature electronically, hence there is a gap in expertise, among ecologically and phylogenetically important taxon,³⁶ which has lead to taxonomic impediment.^{37,38}

In India, not only do we lack an updated checklist of fishes, but also the identification keys which have not been updated after the work of Talwar and Jhingran (1991),³⁹ KC Jayaram(2010)⁴⁰ and Sarma and Mankodi (2017).⁴¹ Also the original descriptions are referred to forever, irrespective of the quality of the paper. Making descriptive taxonomic literature available online is still a major task to promote quality in taxonomy, the unavailability of which impacts the taxonomic process, and often leads to erroneous results and phylogenetic assumptions.³⁶

Academics are currently researching and utilising cutting-edge identifying techniques as a result of these difficulties. The application of DNA technologies for fish identification as a potent substitute tool has overcome the limitations of morphology-based identification approaches and the lack of local fish taxonomists.¹⁹

Dna Barcoding

Paul Hebert (2003) created the idea of DNA barcoding as a molecular identification tool, and it is now a frequently used approach for species identification even by non-specialists. Cytochrome oxidase subunit 1(COI), a mitochondrial DNA gene

utilised as a universal bio-identifying system for an animal, is typically used as a short, standardised nucleotide sequence of DNA for the identification of fish in the process of DNA barcoding⁴² Near the 5'end of the mitochondrial gene cytochrome c oxidase subunit 1(COI) is a 648 base pair segment known as the animal barcode region.

The idea behind this method is that even within the same species, some components of an organism's DNA would vary individually. Finding these components at the species level was the very first task for the scientists who created this method. Geographic isolation causes some populations to stop sharing genetic material, and over time, separate gene pools evolve. These sub-populations maintain morphological similarity but diverge genetically, making them unable to mate and create offspring. These species are known as cryptic species. Because of this speciation, the morphological study of these populations can become questionable as we can't be exactly sure. Such situations can be easily dealt with the molecular characterization method of DNA barcoding.43,44 Thus, it has now become a widely accepted and essential method for proper identification of species on a molecular level.

WHY COI?

This method is employed because mitochondrial DNA has unique properties, such as maternal inheritance, a high copy number per cell, a lack of recombination, a lack of introns, and a greater nucleotide substitution rate, which cause variations between species to rapidly accumulate. Due to COI's low mutation rate compared to other mitochondrial genes in animals, which facilitates its recovery using polymerase chain reaction, it was also chosen as the barcode marker.⁴⁵ A comparative analysis of three mitochondrial genes i.e16S rRNA, cytochrome b, and cytochrome oxidase subunit I (COI) revealed that cyt b and COI are appropriate for clear identification of fishes whereas the 16S rRNA fails to discriminate closely related fish species.⁴⁶

Bar Coding of Fishes Globally and in India

DNA barcoding finds immense application and success in fisheries and furthers the results of conventional morphometry. It is now well established and practiced all across the world. The costs involved in performing the experiment were very expensive in the past but are declining with advancements in technology.⁴⁷

It is commonly believed that taxonomy and barcoding compete with one another for financing, but in reality, entities other than those supporting taxonomic work fund barcoding programmes. Therefore, bar coding would not in any way compete with traditional taxonomy, and the money spent on bar coding is also used to collect and preserve specimens, which are crucial for taxonomy. Therefore, the DNA barcoding programme has the potential to significantly increase fresh financing for museums, herbaria, and individual taxonomy labs rather than reducing support for taxonomy.⁴⁸

Many workers have successfully tested the methodology of barcoding in not only identifying the species but also in the discovery of new species, monitoring of fisheries quotas, correct identification of fisheries products in market, keeping a check on trade of endangered species and identification of cryptic species.^{49,50,51,52,53} Molecular characterization also assists in confirming the absence or presence of a species in given area.⁵⁴

In India, many researchers examined lchthyofaunal diversity using DNA barcoding as a molecular appliance both for marine as well as freshwater fishes. One of the earliest work done on bar coding of fishes in India was by Lakra *et al.*(2011)⁵⁵ for validating the application of bar coding.

Meanwhile many other researchers working on barcoding have confirmed its role in correct identification of fishes^{56,57,58,59,60} and also that barcoding enhances the global data base for quick identification of fishes, validates the checklist of fish fauna of the area, identifies invasive species and helps in formulation of conservation strategies.^{61,62}

Jammu & Kashmir at a Glance

Jammu & Kashmir, the northernmost state in India, is located between 32.17" and 36.58" north latitude and between 73.26" and 80.30" east to west longitude. Due to its uneven topography, the weather in Jammu & Kashmir varies drastically.

Jammu Region

Although the region is sufficiently far west compared to the region's regular 40 to 50 mm (1.6 to 2 inches) of rainfall per month between January and March, the southern regions around Jammu typically have a monsoonal climate. Jammu town can see monthly extremes of rainfall of up to 650 millimetres in August and July, while temperatures in the warmer seasons can exceed 104°F.By early October, conditions are cool and incredibly dry, with little rain and temperatures of about 29 °C (84 °F). By September end, rainfall decreases.

Kashmir Region

The region of Kashmir is renowned for its meadows, lakes, and springs. The earliest records of the area reveal that there was once a sizable lake in the valley, which was encircled by snow-covered mountains. It is thought that Kashmir Valley was once affected by earthquakes that it split apart the mountain wall near Baramulla, letting the water from Satisar Lake pour out and leaving behind karewas, or lacustrine mud, on the mountain edges. For hundreds of millions of years, Kashmir Valley was submerged beneath the Tethya Sea, and the valley's present-day tall sedimentary rock hills were originally submerged in water. The circular but erratic Valley of Kashmir was created in this way. There are many bodies of water in this area, which has a temperate climate.⁶³

Water Bodies and Fish Fauna of Kashmir Region

 Table 1. Showing list of lentic and lotic water bodies in Kashmir region

 WATER BODIES OF KASHMIR⁶⁴

1Dal lake 2Anchar. 3Hokersar 4Nambli Narkara. 5Wular. 6Ajas Wetlands.

15 Sheikhsar 16 Waskursar 17Manasbal Lake 18Vethnar Lake 19Ratan sar 20Gaditar Lake 29 Jhelum River 30 Neelum River 31 Lidder 32 Rambi 33 Sind river 34 Veshaw

7Hygham.	21Sheshnag Lake
8Tarsar Lake	22Marsar Lake
9Mirgund	23Haigam Jeel
10Vishansar	24 Krishansar
11Satsar	25Nundkol Lake
12Nilnag Lake	26 Gadsar
13Kounsarnar	27 Demansar
14Didufnag Lake	28 Gangbal lake

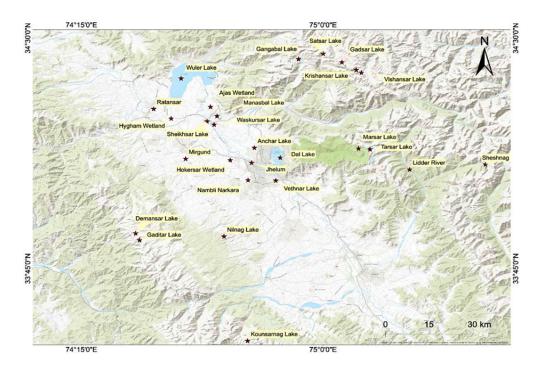


Fig 1: Displaying the GIS-marked Lentic and Lotic water bodies in the Kashmir region.

Fish Fauna of Kashmir Region

The importance of the fish fauna has substantially increased since the endeavour of Haeckel in 1838,⁶⁵ when he published "Fischeaus Caschmir" and thereafter various renowned ichthyologists have come up with very ingenious work like Day (1876),⁶⁶ Silas (1960),⁶⁷ Das and Subla (1964),⁶⁸ Das and Nath(1965),⁶⁹ Yousuf (1996),⁷⁰

Kullander *et al.*(1999),⁷¹ Enderlin and Yousuf (1999),⁷² Balkhi (2007).⁷³

Following table is based on compilation of most recent data obtained from details provided by numerous workers regarding the region's fish fauna as determined by conventional morphometry.

WATERBODY/REFERENCE	Lotic	Lentic	FISHES FAUNA REPORTED
1 River Jhelum a. khan and ali (2013) ⁷⁴	+		ORDER:CYPRINIFORMES FAMILY:CYPRINIDAE 1 Schizothorax curvifrons 2 Schizothorax esocinus

Table 2:	Status of	of fish	fauna	in Ka	ashmir	region.

b. Jan <i>et al.</i> (2015)⁵	÷	3 Schizothorax plagiostomus 4 Schizothorax labiatus 5 Schizothorax niger 6 Cyprinus carpio ORDER:CYPRINIFORMES FAMILY:CYPRINIDAE 1 Schizothorax plagiostomus 2 Schizothorax curvifrons 3 Schizothorax esocinus 4 Schizothorax labiatus 5 Cyprinus carpio ORDER:SALMONIFORMES FAMILY:SALMONIFORMES FAMILY:SALMONIDAE 6 Salmo trutta fario
c. Ahmed <i>et al.</i> (2017) ⁷⁶	÷	7 Salmo gairdneri RIVER:JHELUM ORDER :CYPRINIFORMES FAMILY :CYPRINIDAE 1 Schizothorax Esocinus 2.Schizothorax Plagiostomus 3 Schizothorax Labiatus 4 Schizothorax Curvifrons 5.Schizothorax Niger 6 Cyrinus Carpio Communis 7.Cyprinus Carpio Specularis FAMILY:NEMACHEILIDAE 8 Crossochelius Diplochilus 9 Triplophysia Kashmirensis 10.Triplophysa Marmorata
2. Anchar Lake- Bashir <i>et al.</i> (2016) ⁷⁷	÷	ORDER :SALMONIFORMES FAMILY:SALMONIDAE <i>1 Salmo trutta fario</i> ORDER:CYPRINIFORMES FAMILY:CYPRINIDAE 2 S. plagiostomus 3 Schizothorax ecocinus 4 S.labiatus 5 S. niger 6 S. richardsoni 7 S. curvifrons 8 Crossocheilus diaplochilus 9 Bangana diplostoma 10 Cypinus carpio communis 11 Cyprinus carpio specularis 12 Puntius conchonius 13 Carassius carassius FAMILY:NEMACHEILIDAE 14 Tritlophysa kashmirensis ORDER:CYPRINIDONTIFORMES FAMILY:POECILIIDAE <i>15 Gambusia affinis</i>
3.Wular Wetland- a.Brraich and malik (2016) ⁷⁸	+	ORDER :CYPRINIFORMES FAMILY:CYPRINIDAE

770

	1Schizothorax labiatus
	2 Schizothorax esocinus
	3 Cyprinuscarpio var. communis
	4 Schizothorax micropogon
	5 Cyprinuscarpio var. specularis
	6 Ctenopharyngodon idella
	7 Schizothorax richardsonii
	8 Schizothorax niger
	9 Carassius carassius
	10 Schizothorax curvifrons
	11 Crossocheilus latius
	FAMILY:NEMACHEILIDAE
	-
-1	12Triplophysa marmorata
	ORDER:CYPRINIFORMES
et al.(2016) ⁷⁹	+ FAMILY:CYPRINIDAE
	1Cyprinus carpio specularis
	2 Cyprinus carpio communis
	3 Carassius carassius
	4 Schizothorax niger
	5 Schizothorax esocinus
	6 Schizothorax curvifrons
	7 Schizothorax labiatus
	8 Schizothorax plagiostomus
	9 Crossochelius diplochilus
	10 Puntius conchonius
	FAMILY: COBITIDAE
	11 Botia birdi
	FAMILY:NEMACHEILIDAE
	12 Triplophysa kashmirensis
	13 Triplophysa marmorata
	ORDER:CYPRINODONTIFORMES
	FAMILY:POECILIIDAE
	14Gambusia affinis
	ORDER:SILURIFORMES
	FAMILY:SISORIDAE
	15 Glyptothorax kashmirensis
	16 Glyptothorax pectinoptrus
<i>al.</i> (2018) ⁸⁰	ORDER:CYPRINIFORMES
	+ FAMILY:CYPRINIDAE
	1 Cyprinus carpio var. communis
	2Cyprinus carpio var. specularis
	3Carassius carassius
	4Schizothorax niger
	5Schizothorax esocinus
	6Schizothorax curvifrons
	7Crossocheilus diplocheilus
	8Puntius conchonius
	FAMILY:NEMACHEILIDAE
	9Triplophysa spp
et al (2017) ⁷⁶	
<i>et al.</i> (2017) ⁷⁶	+ FAMILY :CYPRINIDAE

b Wular Lake-Rumysa *et al.*(2016)⁷⁹

c-Qadri et al.(2018)80

4.Dal lake a-Ahmed *et al.*(2017)⁷⁶

			1 <i>Cyrinus Carpio Communis</i> 2. <i>Cyprinus Carpio Specularis</i> 3 <i>SchizothoraxCurvifrons</i> 4. <i>Schizothorax Niger</i> 5. <i>CrossocheilusDiplochilus</i> 6. CarassiusCarassius ORDER:CYPRINODONTIFORMES FAMILY:POECILIIDAE 7. <i>PuntiusConchonius</i> 8. <i>GambusiaHolbrooki</i> FAMILY:BOTIDAE 9. <i>BotiaBirdi</i>
5.Hokersar Wetland-			ORDER:CYPRINIFORMES
Mushtaq <i>et al.</i> (2019) ⁸¹		+	FAMILY:CYPRINIDAE
			1Cyprinus Carpio. Communis 2 Cyprinus Carpio Specularis 3 Schizothorax Niger
6.River Viashaw-			ORDER :CYPRINIFORMES
a-Hamid and Singh(2019)82	+		FAMILY:CYPRINIDAE
a-Hamid and Singh(2019)	т		1 Schizothorax plagiostomas
			2 Schizothorax curvifrons
			3 Schizothorax esocinus
			4 Schizothorax richardsonii
			5 Triplophysa kashmirensis
			6.Crossocheilus diplochilus
b-Rashid and singh (2020) ⁸³			ORDER:CYPRINIFORMES
b-rtashia ana singir (2020)	+		FAMILY:CYPRINIDAE
			1 Schizothorax plagiostomus
			2 Schizothorax labiatus
			3 Schizothorax esocinus
			4 Schizothorax curvifrons
			5 Cyprinus carpio communis
			FAMILY:NEMACHEILIDAE
			6 Triplophysa kashmirensis
			7 Triplophysa marmorata
			ORDER: SILURIFORMES
			FAMILY: SISORIDAE
			8 Glyptosternon reticulatum

Table 3: Summary of fish species found in Kashmir region(based on compilation of data of table 2)

ORDER	FAMILY
1.CYPRINIFORMES	CYPRINIDAE 1. Bangana diplostoma 2. Cyprinus carpio 3. Crossochelius Diplochilus 4. Crossocheilus latius 5. CarassiusCarassius

	6. Ctenopharyngodon idella
	7. Puntius conchonius
	8. Schizothorax curvifrons
	9. Schizothorax esocinus
	10. Schizothorax plagiostomus
	11. Schizothorax micropogon
	12. Schizothorax labiatus
	13. Schizothorax niger
	14. Schizothorax richardsonii
	15. Triplophysa kashmirensis
	COBITIDAE
	1. Botia birdi
	NEMACHEILIDAE
	1. BotiaBirdi
	2. Crossochelius Diplochilus
	3. Crossocheilus latius
	4. GambusiaHolbrooki
	5. PuntiusConchonius
	6. Triplophysa kashmirensis
	7. Triplophysa marmorata
2.CYPRINIDONTIFORMES	POECILIIDAE
	1. Gambusia affinis
3.SALMONIFORMES	SALMONIDAE
	1. Salmo trutta fario
	2. Salmo gairdneri
	NEMACHEILIDAE
	1. Triplophysa kashmirensis
	2. Triplophysa marmorata
4. SILURIFORMES	SISORIDAE
	1. Glyptothorax kashmirensis
	2. Glyptothorax pectinoptrus
	3. Glyptosternon reticulatum

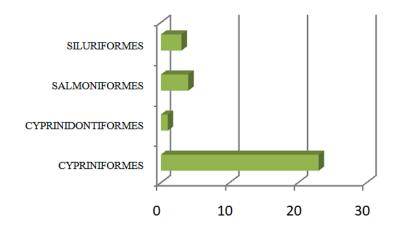
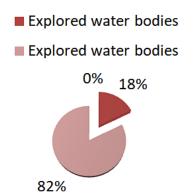
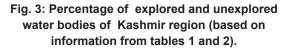


Fig. 2:Percentage contribution of different orders to fish diversity of Kashmir region (according to table 3's data)

Analysis of the data in table 2 demonstrates that only a small portion of the total number of water bodies existent (given in table 1) have been extensively studied, leaving the majority of water bodies undiscovered.





Current Status of Barcoding Integrated with Conventional Morphometry in the Valley.

Analysis of the data (table 2) reveals that different researchers have identified different species in the same body of water, and also that there is not a significant temporal gap between those reports, therefore taxonomic ambiguity must have been a major factor in the inconsistent results reported, as many of the species native to the region are challenging to identify using conventional morphometry. Like Identification of species of genus *Schizothorax* and *Tryplophysa* through conventional methodology can some times lead to errornous results.

The morphology of the genus *Schizothorax* is strikingly similar, making it challenging to distinguish between species based on morphological characteristics. Lately collaboration of barcoding with conventional morphometry has been adopted and tested. After performing morphometric characterization to see if barcoding can aid in accurate species identification in fishes, researchers DNA-barcoded *schizothorax* species from the Neelum and Jhelum rivers in Azad Kashmir. The results showed that barcoding is accurate, dependable, and has enormous potential for species identification.⁸⁴ In addition to that a similar study on five different species of *Schizothorax* validated the role of cytochrome oxidase I in species delineation in conjunction with morphometric information and also that the Sequence-based phylogenetic analysis reveals different species groups⁸⁵ (Bashir *et al.*, 2015).

This integrated approach was also used to characterise two significant fish species from the Kashmir valley, Triplophysa marmorata and T. kashmirensis. Due to the inadequate original descriptions and the dearth of positive reviews, it is difficult to distinguish between these two species. A morphometric and molecular analysis was carried out with this in mind. Investigation concluded that these two taxonomic Triplophysa taxa should be accepted as valid based on morphological and mt DNA COI sequence analyses. These findings can help ichthyologists better understand the ichthyofauna of the Kashmir valley and may aid them in developing methods for protecting and managing these lesser-studied native tiny species within their area of distribution²¹

Water Bodies and Fish Fauna of Jammu Region Jammu region with subtropical climate is blessed with a number of lentic and lotic water water bodies offering ample water resources for development of fisheries

2.Pargwal Wetland13.Sangral Wetland1	 Chenab River Tawi River Ravi River Poonch River
--------------------------------------	--

WATER BODIES OF JAMMU⁶⁴

Fish Fauna of Jammu Region

Icthyofauna of the Jammu region was intensively investigated for the first time by Das and Nath (1965,1966)⁸⁶ eventually many workers have reported fish fauna from the region like Das and Nath (1971),⁸⁷ Malhotra *et al.* (1975)⁸⁸ Joshi *et al.* (1978),⁸⁹Tilak (1971)⁹⁰ Dutta and Malhotra(1984),⁹¹ Jyoti *et al.* (2006)⁹² and Balkhi (2007).⁷³ Following table is based on a collection of recent information on fish diversity provided by multiple workers for various lentic and lotic water bodies in the Jammu region, as determined by conventional morphometry.

WATERBODY/ REFERENCE	LOTIC	LENTIC	FISHES FOUND
-	+		PISHES FOUND ORDER:CYPRINIFORMES FAMILY:CYPRINIDAE 1. Schizothorax plagiostomus 2. S. labiatus 3. Tor putitora 4. T.tor 5. Tor khudree 6. Crossocheilus latius 7. Garra gotyla 8. G.lamta 9.Barilius vagra 10. B.bendelisis 11. Labeo rohita 12. Labeo bata 13. Puntius conchonius 14. P. sophore 15. P. ticto 16. Schizothorax richardsoni 17. Cyprinus carpio 18. Cirrhinus reba FAMILY:NEMACHEILIDAE 19. Nemacheilus botia FAMILY:COBITIDAE 20.Botia dayi ORDER:SYNBRANCHIFORMES FAMILY:MASTACEMBELIDAE 21 Mastacembalus armatus 22 Macrognathus pancalus ORDER:SILURIFORMES FAMILY:SISORIDAE 23. Glyptothorax botium
			23. Gryptotriorax bottom 24. G. pectinopterum 25. Glyptosternum maculatum 26 Bagarius bagarius FAMILY:SILURIDAE 27 Wallago attu
			FAMILY:BAGRIDAE 28. <i>Mystus seenghala</i> 39. <i>M. bleekeri</i> 30 <i>Mystus cavasius</i> ORDER:BELONIFORMES
b.River Chenab Kishtwar district. Bhutyal and Langer(2015) ⁹⁴	+		FAMILY:BELONIDAE 31 Xenentodon cancila ORDER:SALMONIFORMES FAMILY:SALMONIDAE. 1 Oncorhynchus mykiss ORDER:SILURIFORMES FAMILY : SISORIDAE

Table 5: Status of fish fauna reported from the Jammu region

2.River Tawi Gandotra et. al (2017)⁹⁵ +

+

3.River Basantar Sharma and Dutta (2012)⁹⁶ 2 Glyptosternum reticulatum **ORDER : CYPRINIFORMES** FAMILY:CYPRINIDAE 3 Schizothorax sp. 4 Cyprinus carpio 5.Schizothorax richardsonii **ORDER : CYPRINIFORMES** FAMILY:CYPRINIDAE 1 Garra gotyla 2 Schizothorax richardsonii 3 Labeo boga 4 Tor putitora 5 Barilius vagra 6 Puntius ticto 7 Puntius conchonius 8 Aspidoparia morar 9 Crossocheilus latius 10 Barilius bendelisis 11 Schizothorax richardsonii 12 Labeo bata 13 Puntius sophore **ORDER : BELONIFORMES** FAMILY:NEMACHEILIDAE 14 Schistura montanus 15 Nemacheilus botia ORDER:MASTACEMBELIFORMES FAMILY:MASTACEMBELIDAE 16 Mastacembelus pancalus 17 Mastacembelus armatus **ORDER:PERCIFORMES** CHANNIDAE 18 Channa punctatus 19 Channa striatus **ORDER: SILURIFORMES** SISORIDAE 20 Bagarius yarrelli BAGRIDAE 21 Mystus seenghala **ORDER : OSTEOGLOSSIFORMES** FAMILY : NOTOPTERIDAE 1Notopterus notopterus **ORDER : CYPRINIFORMES** FAMILY:CYPRINIDAE 2 Catla catla 3. Cirrihinus mrigala 4. Cirrihinus reba 5. Labeo gonius 6. Labeo rohita 7. Labeo calbasu 8. Puntius ticto 9. Puntius sophore 10. Puntius sarana 11. Puntius chola 12. Tor tor 13.Aspidopario morar 14. Barilius vagra 15. Danio devario 16. Rasbora rasbora 17. Crossocheilus latius diplochilus 18. Garra gotyla FAMILY:COBITIDAE 19. Botia davi ORDER:SILURIFORMES FAMILY:BAGRIDAE 20. Mystus bleekeri 21. Mystus vittatus 22. Aorichthys seenghala 23. Rita rita

4.Ornamental Fishes of Jammu Region a.Vohra *et al.* (2013)⁹⁷

b.Arif et al. (2019)98

+

FAMILY:SILURIDAE 24. Ompak bimaculatus 25. Wallago attu FAMILY:SCHILBEIDAE 26.Clupisoma garua FAMILY:SISORIDAE 27. Bagarius bagarius 28. Gagata cenia 29. Glyptothorax stoliczkae **ORDER : PERCIFORMES** FAMILY : NANDIDAE 30. Badis badis FAMILY:CHANNIDAE 31. Channa punctatus 32. Channa marulius 33. Channa orientalis **ORDER : SYNBRACHIFORMES** FAMILY: MASTACEMBELIDAE 34. Mastacembelus armatus 35. Macrograthus pancalus ORDER:CYPRINIFORMES FAMILY:CYPRINIDAE 1 Danio devario 2 Danio rerio 3 Chela bacaila 4 Esomus danricus 5 Rasbora rasbora 6 Puntius spp 7 Barilius vagra 8 Osterobrama cotia 9 Aspidoparia morar FAMILY: BOTIIDAE 10 Botia davi FAMILY:NEMACHEILIDAE 11 Noemachilus botia FAMILY:COBITIDAE 12 Lepidocephalichthys guntea ORDER:SILURIFORMES FAMILY:BAGRIDAE 13 Mystus bleekri FAMILY:HETEROPNEUSTIDAE 14 Heteropneustes fossilis ORDER:SYNBRANCHIFORMES FAMILY: MASTACEMBELIDAE 15 Macrognathus aculeate 16 Mastacembellus sps. 17 Mastacembellus armatus 18 Mastacembellus pancalus ORDER: ANABANTIFORMES FAMILY:OSPHRONEMIDAE 19 Trichogaster fasciatus **ORDER : BELONIFORMES** FAMILY:BELONIDAE 20 Xenentodon cancilla 21 Aspidoparia morar **ORDER : CYPRINIFORMES** FAMILY:CYPRINIDAE 1Salmostoma bacaila

5.Erstwhile Udhampur District Dutta (2015)⁹⁹

+

2 Salmostoma panjabensis 3 Aspidoparia morar 4 Barilius vagra 5 Barilius bendelisis 6 Rasbora rasbora 7 Esomus danricus 8 Danio devario 9 Chela cahius 10 C.laubuca 11Tor tor 12 T. putitora 13 Puntius sophore 14 P. chola 15 P. ticto 16 P. conchonius 17 P.sarana 18 Crossocheilus latius FAMILY:NEMACHEILIDAE 19 Nemacheilus botia **ORDER:SILURIFORMES** FAMILY : BAGRIDAE 20 Mystus seenghala 21 Mystus bleekeri FAMILY:HETEROPNEUSTIDAE 22 Heteropneustes fossilis **ORDER:BELONIFORMES** FAMILY:BELONIDAE 23 Xenentodon cancilia ORDER:SYNBRACHIFORMES FAMILY:MASTACEMBELIDAE 24 Mastacembelus armatus 25 Macrognathus pancalus ORDER: ANABANTIFORMES FAMILY : CHANNIDAE 26 Channa punctatus 27 C. striatus 28 C.maruilius FAMILY:OSPHRONEMIDAE 29 Trichogaster fasciatus **ORDER:PERCIFORMES** FAMILY:BADIDAE 30 Badis badis **ORDER: CYPRINIFORMES** FAMILY : CYPRINIDAE 1 Hypothalmichthys molitrix 2 Salmostoma bacaila 3 Aspidoparia morar 4 Barilius vagra vagra 5 B. bendelisis 6 B. shacra 7 B. modestus 8 B. radiolatus 9. Esomus danricus 10 Danio devario 11Brachydanio. rerio 12 Rasbora. rasbora 13 Amblypharyngodon. mola 14 Ctenopharyngodon idellus 15 Cyprinus carpio communis 16 C. carpio specularis 17 Tor. tor 18. T. putitora 19 . Catla catla 20 Osteobrama. cotio cotio 21 Puntius.sarana sarana 22 Puntius, conchonius 23 P. terio 24 P. ticto 25 P. chola 26 P. sophore 27 Cirrhinus mrigala 28 C reba 29 Labeo bata 30. Labeo calbasu

778

31. L. dero 32. L. dyocheilus dyocheilus 33. L. pangusia 34. L. rohita 35. Schizothrax. richardsonii 36. Schizothorichthys. progastus 37. S. esocinus 38. S. curvifrons 39. Crossoscheilus. latius diplocheilus 40. Garra. gotyla gotyla 41. Garra. lamta 42. Nemacheilus. corica 43. Acanthocobitis.botia 44. Schistura. punjabensis 45. S.montanus 46. Triplophysa. yasinensis 47. Botia. almorhae 48. B. birdi 49. B. lohachata 50. B. dario 51. Lepidocephalus. guntea **ORDER: SILURIFORMES** FAMILY : BAGRIDAE 52. Mystus. bleekeri 53. M. cavasius 54. M. vittatus 55. Aorichthys. seenghala FAMILY :SILURIDAE 56. Ompok. bimaculatus 57. O. pabda 58. Wallago. attu FAMILY : SCHILBEIDAE 59. Eutropiichthys. vacha FAMILY : AMBLYCIPITIDAE 60. Amblyceps.mangois FAMILY :SISORIDAE 61. Bagarius. bagarius 62.Glytosternon. reticulatum 63. Glyptothorax. pectinopterus 64. G. indicus 65. G. telchitta telchitta 66. G. cavia 67. G. Kashmirensis 68. G. punjabensis FAMILY:HETEROPNEUSTIDAE 69. Heteropneustes. fossilis **ORDER : BELONIFORMES** FAMILY:BELONIDAE. 70. Xenentodon. cancila **ORDER : SYNBRANCHIFORMES** FAMILY: MASTACEMBELIDAE 71. Macroganthus. pancalus 72. Mastacembelus. armatus **ORDER : PERCIFORMES** FAMILY: BELONTIDAE 73. Colisa fasciatus FAMILY :CHANNIDAE 74. Channa. Orientalis 75. C. punctatus **ORDER :SALMONIFORMES** FAMILY : SALMONIDAE 76. Salmo trutta fario

6.River Ujh, an important tributary of the river Ravi. Rathore and Dutta (2015)¹⁰⁰

+

7.Chadwal Stream Khajuria *et al.*(2015)¹⁰¹ **ORDER : CYPRINIFORMES** FAMILY: CYPRINIDAE 1. Salmostoma bacaila 2. Salmostoma panjabiensis 3. Aspidoparia morar 4. Barilius vagra vagra 5. B. bendelisis 6. Rasbora rasbora 7. Esomus danricus 8. Danio devario 9. Tor tor 10. T. putitora 11. Puntius sophore 12. P. chola 13. P. ticto 14 P. conchonius 15. Cirrhinus mrigala 16. C. reba 17. Labeo dero 18. L. dyocheilus 19. L. pangusia 20. Catla catla 21. Crossocheilus latius diplocheilus 22. Garra lamta 23. G. gotyla FAMILY: BALITORIDAE 24. Acanthocobitis botia FAMILY:COBITIDAE 25. Botia almohare 26. Botia birdi 27. Lepidocephalichthys guntea **ORDER : SILURIFORMES** FAMILY : BAGRIDAE 28. Aorichthys seenghala 29 Mystus bleekeri 30 Mystus Vittatus FAMILY : SILURIDAE 31 Ompok bimaculatus 32 Wallago attu FAMILY: AMBLYCIPITIDAE 33. Amblyceps mangois FAMILY:SISORIDAE 34. Bagarius bagarius 35. Glyptothorax pectinopterus 36. G. stoliczkae 37. G. telechitta telechitta **ORDER:BELONIFORMES** FAMILY:BELONIDAE 38. Xenentodon cancilia ORDER:SYNBRANCHIFORMES FAMILY:MASTACEMBELIDAE 39.Mastacembelus armatus 40.Macrognathus pancalus ORDER:PERCIFORMES FAMILY:CHANNOIDAE 41. Channa punctatus 42. C. orientalis **ORDER : CYPRINIFORMES** FAMILY:CYPRINIDAE 1 Labeo boga 2 Puntius ticto 3 Labeo dero 4 Labeo calbasu 5 Labeo rohita 6 Danio devario 7 Crossocheilus latius 8 Puntius sophore 9 Puntius sarana 10 Chela bacaila

8.Sunderbani(stream) Gandotra and Sharma (2015)¹⁰²

+

+

9. Lotic waterbodies of r.s.pura Tehsil Kour *et al.* (2015)¹⁰³ 11 Barilius bendelesis 11 Barilius vagra 12 Garra gotyla gotyla 18 Tor putitora FAMILY:NEMACHEILIDAE 19 Nemacheilus botia **ORDER : BELONIFORMES** FAMILY:BELONIDAE 20 Xenentodon cancila **ORDER: ANABANTIFORMES** FAMILY:CHANNIDAE 21Channa punctatus **ORDER:CYPRINIFORMES** FAMILY: CYPRINIDAE 1 Schizothorax richardsonii 2 Tor putitora 3 Garra gotyla 4 Labeo boga 5 Labeo bata 6 Labeo dero 7 Crossocheilus latius 8 Puntius conchonius 9 Puntius sophore 10 Puntius ticto 11 Barilius bendelisis 12 Barilius vagra **ORDER: SILURIFORMES** FAMILY :SISORDAE 13 Glyptothorax pectinopterus **ORDER: CYPRINIFORMES** FAMILY:CYPRINIDAE 1. Amblypharyngodon mola 2. Barilius bendelisis 3. Garra gotyla gotyla 4. Osteobrama cotio 5. Puntius sophore 6. Puntius ticto7. Puntius conchonius 8. Salmostoma bacaila 9. Aspidoparia morar 10. Danio devario 11. Chela laubucca 12. Barilius vagra 13. Esomus danricus 14. Labeo boga FAMILY:COBITIDAE 15 Lepidocephalichthys guntea 16 Noemacheilus botia **ORDER:SILURIFORMES** FAMILY:BAGRIDAE 17. Mystus bleekeri 18. Mystus seenghala 19. Mystus vittatus FAMILY:SILURIDAE 20. Wallago attu FAMILY:SCHILBEIDAE 21. Pseudoeutropius athernioides **ORDER:PERCIFORMES** FAMILY: AMBASSIDAE 22. Ambassis nama **ORDER : BELONIFORMES** FAMILY:BELONIDAE 23. Xenentodon cancila **ORDER : OPHIOCEPHALIFORMES** FAMILY:OPHIOCEPHALIDAE 24. Channa punctatus

10.Wajoo nullah (an important tributary of the river) Dutta (2016)¹⁰⁴ +

ORDER:MASTACEMBELIFORMES FAMILY:MASTACEMBELIDAE 25. Mastacembelus pancalus **ORDER: OSTEOGLOSSIFORMES** FAMILY:NOTOPTERIDAE 1 Notopterus notopterus 2 Chitala chitala **ORDER: CLUPEFORMES** FAMILY: CLUPEIDAE 3. Gudusia chapra **ORDER : CYPRINIFORMES** FAMILY:CYPRINIDAE 4 Securicula gora 5 Salmostoma bacaila 6. S.punjabensis 7. Aspidoparia morar 8. Barilius bendelisis 9. B.vagra vagra 10. B.modestus 11. Danio.devario 12. Esomus danricus 13. Rasbora rasbora 14. Amblypharyngodonmola 15. Chela laubuca 16. Chela cachius 17. Cyprinus carpio communis 18. Tor tor 19. Tor putitora 20. Puntiusticto 21. P.sophore 22. P.saranasarana 23. P.conchonius 24. Labeo.bata 25. Labeo.dero 26. L.dyocheilus dyocheilus 27. L.gonius 28. L.calbasu 29. L.pangusia 30. L.boggut 31. Cirrhinus.mrigala 32. C.reba 33. Osteobrama cotio cotio 34. Crossocheilus.latitus diplocheilus 35. Garra gotyla gotyla 36. G.lamta FAMILY: BALITORIDAE 37. Acanthocobitis botia FAMILY: COBITIDAE 38. Botia almorhae 39. B. lohachata 40. Lepidocephalus guntea **ORDER: SILURIFORMES** FAMILY: BAGRIDAE 41. Rita rita 42. M.vittatus 43. M.bleekeri 44. M.cavasius 45. Aorichthys seenghala FAMILY: SILURIDAE 46.Ompok pabda 47. Wallago attu FAMILY: SCHILBIDAE 48. Pseudeutropius atherinoides FAMILY: AMBLYCPTIDAE 49 Amblyceps mangois FAMILY:SISORIDAE 50. Bagarius bagarius 51. Gagata cenia FAMILY:HETEROPNEUSTES 52 Heteropneustes fossilis

11.Tehsil mendhar (district- poonch) Hussain and Dutta (2016)¹⁰⁵

+

+

12.Selum nullah and Aik nullah Khajuria *et al.*(2016)¹⁰⁶

+

ORDER : BELONIFORMES FAMILY: BELONIDAE 53. Xenontodon cancila **ORDER : SYNBRANCHIFORMES** FAMILY :MASTAMCEMBELIDAE 63. Macroganthus pancalus 64. Mastacembelus armatus **ORDER : CYPRINIFORMES** FAMILY : CYPRINIDAE 1 Cyprinus carpio communis 2 Hypophthalmichthys molitrix 3 Ctenopharyngodon idellus 4 Schizothorax richardsonii 5 L. dero 6 L. dyocheilus dyocheilus 7 L. bata 8 Gara gotyla 9 G. lamta 10 Crossocheilus latius diplocheilus 11 Barilius vagra 12 Tor putitora 13 Puntius conchonius FAMILY : BALITORIDAE 14 Schistura prashadi 15. S. punjabensis 16. S. montanus FAMILY :COBITIDAE 17 Botia birdi **ORDER: SILURIFORMES** FAMILY : SISORIDAE 18 Glyptothorax punjabensis FAMILY : SILURIDAE 19 Ompok pabda **ORDER : SYNBRANCHIFORMES** FAMILY : MASTACEMBELIDAE 20 Mastacembelus armatus **ORDER : PERCIFORMES** FAMILY:CHANNIDAE 21 Channa orientalis **ORDER : CYPRINIFORMES** FAMILY:CYPRINIDAE 1 Puntius conchonius 2 Puntius sophore 3 Garra gotyla gotlya 4 Labeo boga 5 Labeo dero 6 Labeo calbasu 7 Cirrhinus reba 8 Crossocheilus latius 9 Barilius vagra 10 Danio devario FAMILY:COBITIDAE 11 Rasbora rasbora 12 Botia dayi 13 Lepidochephalus thermalis 14 Lepidochephalus guntea **ORDER:SILURIFORMES** FAMILY:SILURIDAE 15 Heteropnuestes fossilis FAMILY:BAGRIDAE 16 Mystus bleekeri FAMILY:SISORIDAE

13. Ichthyofauna of Rajouri district. Nisa *et al.* (2020)¹⁰⁷

+

+

14River Ravi Dutta(2021)¹⁰⁸ 17 Gagata gagata ORDER:SYNBRANCHIFORMES FAMILY:MASTACEMBALIDAE 18 Mastacembalus armatus **ORDER : OPHIOCEPHALIFORMES** FAMILY:OPHIOCEPHALIDAE 19 Channa punctatus **ORDER:BELONIFORMES** FAMILY:BELONIDAE 20 Xenentodon cancila ORDER:CYPRINIFORMES. FAMILY: DANIONIDAE Barilius vagra FAMILY:NEMACHEILIDAE. 2 Triplophysa sp. FAMILY :CYPRINIDAE 3 Cirrhinus mrigala 4 Cyprinus carpio 5 Garra gotyla 6 Garra lamta 7 Labeo bata 8 Labeo boga 9 Bangana dero 10 Labeo rohita 11Puntius sophore 12 Pethia ticto 13 Shizothorax richardsoni 14 Tor putitora 15 Tor tor ORDER:SILURIFORMES. FAMILY:SISORIDAE 16 Glyptothorax pectinopterus **ORDER: OSTEOGLOSSIFORMES** FAMILY: NOTOPTERIDAE 1. Notopterus notopterus 2. Chilata chitala **ORDER: CLUPEIFORMES** FAMILY: CLUPEIDAE 3. Gudusia chapra **ORDER: CYPRINIFORMES** FAMILY:CYPRINIDAE 4. Salmophasia bacaila 5. Salmophasia phulo 6. Salmophasia Punjabensis 7. Securicula gora 8. Asidoparia morar 9. Barilius vagra vagra 10. B. barila 11. B. modestus 1 2. B. radiolatus Gunther 13. B. bendelisis 14. Raiamas bola 15. Chela cachius 16. Chela laubuca 17. Esomus danricus 18. Danio. devario 19. Rasbora daniconius 20. Amblypharyngodon. mola 21. Cyprinus. carpio communis 22. Cyprinus. carpio specularis 23. Tor. tor 24. Tor. putitora 25. Osteobrama. cotio cotio 26. Puntius. sarana sarana 27. P. conchonius 28. P. terio 29. P. ticto 30. P. chola 31. P. sophore 32. Cirrhinus mirgala 33. Cirrhinus reba 34. Catla catla

35. Labeo bata 36. L. boga 37. L. calbasu 38. L. dero 39. L. dyocheilus 40. L. gonius 41. L. pangusia 42. L. rohita 43. L. lippus 44. Schizothorax. richardsonii 45. Crossocheilus. latius diplocheilus 46. C. latius punjabensis 47. Garra. gotyla gotyla 48. G. lamta FAMILY: BALITORIDAE 49. Nemacheilus corica 50. Acanthocobitis botia 51. Schistura prashadi 52. S. montanus 53. S. punjabensis FAMILY: COBITIDIAE 54. Botia almorhae 55. Botia birdi 56. Botia lohachata 57. Lepidocephalus. guntea **ORDER: SILURIFORMES** FAMILY: BAGRIDAE 58. Rita rita 59. Mystus bleekeri 60. M. cavasius 61. M. vittatus 62. M. tengara 63. Aorichthys seenghala 64. A. aor FAMILY: SILURIDAE 65. Ompok pabda 66. Wallago attu FAMILY: SCHILBIDAE 67. Ailia punctata 68. Neotropius atherinoides 69. Clupisomagarua 70. Clupisomanazri 71. Eutropiichthysmurius 72. E. vacha FAMILY: AMBLYCIPITIDAE 73. Amblyceps mangois FAMILY:SISORIDAE 74. Bagarius. bagarius 75. Gagata. cenia 76. Glyptosternum reticulatum 77. Glyptothorax cavia 78. G. conirostreconirostre 79. G. pectinopterus 80. G. stoliczkae 81. G. telchitta FAMILY: CLARIIDAE 82. Heteropneustes fossilis 83. Clarius batrachus **ORDER: SALMONIFORMES** FAMILY: SALMONIDAE 84.Salmo trutta fario FAMILY: BELONIDAE 85. Xenentodon. cancila **ORDER: SYNBRANCHIFORMES** FAMILY: MASTACEMBELIDAE 86. Macroganthus aral 87 M. pancalus 88. Mastacembelus armatus **ORDER: PERCIFORMES** FAMILY: CHANDIDAE

15.Fish Fauna of		 89. Chanda nama 90. Parambassisbaculis 91. P. ranga FAMILY: NANDIDAE 92. Nandus nandus FAMILY: GOBIIDAE 93. Glossogobius giuris FAMILY :CHANNIDAE 94. Channa marulius 95. C. orientalis 96. C. punctatus 97. C. striatus ORDER:CYPRINIFORMES
River Sewa +		FAMILY:CYPRINIDAE
Gupta and		1 Barilius vagra 2 Barilius bendelisis
Dutta (2021) ¹⁰⁹		3 Crossocheilus latius diplocheilus
		4 Tor putitora 5 Cirrhinus reba
		6 Schizothorax richardsonii ORDER:SILURIFORMES
		FAMILY:SISORIDAE
		7 Glyptothorax stoliczkae
		ORDER:SALMONIFORMES
		FAMILY:SALMONIDAE
		8 Salmo trutta fario
16.Mansar-Surinser		ORDER:CHANNIFORMES
Lake (Information Sheet	+	FAMILY:OPHIOCEPHALIDAE
on Ramsar sites)		1.Channa gachua 2.Channa punctatus
		ORDER:CYPRINIFORMES FAMILY:CYPRINIDAE
		3.Cyprinus carpio 4.Danio rerio
		5.Labeo rohita 6.Puntius chonchonius
		7.Rasbora rasbora
		FAMILY:BELONTIIDAE
		8. Trichogaster fasciatus
		FAMILY:MASTACEMBELIDAE
		9.Mastacembelus armatus
17.Gharana wetland		ORDER:CYPRINIFORMES
(Information Sheet	+	FAMILY:CYPRINIDAE
on Ramsar sites)		1.Puntius sophore 2.Puntius ticto 3.Rasbora rasbora
		FAMILY:CHANNIDAE
		4Channa marulius 5Channa orientalis
		6Channa punctatus 7Channa striatus
		FAMILY:BELONTIIDAE
		8Trichogaster fasciatus
		FAMILY:CLARIIDAE
		9Heteropneustes fossilis

According to an analysis of the data in table 5, only a small percentage of the total number of water bodies that are known to exist (as shown in table 4) have undergone comprehensive research, leaving the bulk of water bodies unexplored.

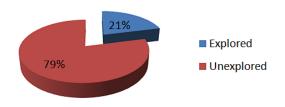


Fig. 4: Showing percentage of explored and unexplored water bodies of Jammu region.

Status of Bar Coding in Jammu Region

The present study, which is based on an examination of past findings, found that several lentic and lotic water bodies in the area has documented about 160 species. However, due to a lack of molecular characterization work, the employment of contemporary methodologies is still in its infancy.

Arif and Gandotra (2017)¹¹⁰ carried out DNA barcoding of ornamental fishes in various water bodies in the Jammu region for the first time, verifying its usage for precise species identification. Analysis of the economic value of the fish fauna of the region reveals that the majority are food fishes, with ornamental fishes accounting for the secondhighest percentage, and the remainder have both food and ornamental fishes as economic status. Since food and forage fish make up the majority of the fish in the area, molecular characterization of the major section of fish diversity remains untouched, placing the data available regarding the current status of fish fauna under uncertainty.

As shown by the data in table 5, the Jammu region is blessed with a rich diversity of fish species, many of which are generally difficult to identify morphometrically. However, bar coding is still in its infancy in this area. Major factors contributing to this research gap include outdated knowledge of current techniques, lack of funding, greater expertise in conventional methodologies, and most importantly, the fact that basic research is being neglected in favour of applied research as taxonomy has taken a backseat over time. Because most of the species lack accurate taxonomic resolution, analyses of the historical record of fish distribution, making temporal comparisons, and tracking the proper phylogeny have all been impeded.

Discussion

Due to the difference in topography, larger number of lotic water bodies, and more favourable climatic conditions, the fish diversity in the Jammu region is greater than that in the Kashmir region. Cypriniformes is the most prevalent order in both Jammu and Kashmir due to their great level of adaptability and capacity to occupy any area. Along with the endemic species of the genus *Schizothorax*, many of the fishes listed above (tables 4 and 5) are exotic species, such as carps, which are not native to the area and were introduced by the state fisheries department.

Anthropogenic Stress, Declining Fish Diversity and Need for Conservational Measures

Fish, which have a heterothermic body temperature, are easily impacted by changes in the physicochemical characteristics of the body of water they reside in.^{111,112} The aquatic ecosystem is being negatively impacted by climate change and anthropogenic factors such as pollution, overfishing, hydropower projects, etc. These factors also cause coral bleaching, the loss of coastal wetland, changes in the distribution and timing of freshwater flow, and a decline in fish diversity.¹¹³

The largest freshwater lake in Asia, Wular Lake, is home to several fish species. However, eutrophication caused by human activity, which alters the water's physicochemical properties and impairs ecological conditions, has caused the extinction of numerous *schizothorax* species that are adapted to clean water.^{79,80} Fish population of *Schizothorax plagiostomus* and *Schizothorax esocinus* in Dal lake has also been affected because of the constantly degrading water quality of the water body.¹¹⁴

The River Jhelum, a significant tributary of the Indus River System that drains through the entire state of Kashmir, is a celebrated river economically and a significant source of water for expanding human population and irrigation. However, the water body's shifting biological conditions have also encouraged the effective colonisation of foreign fish species with exceptional adaptability. The Viashaw River, a left tributary of the Indus River System, is also being affected by illicit mining and overfishing, which is reducing the diversity of fish.^{83,114} Water pollution has affected the fish fauna of Jammu region as well, a comparative analysis of fish fauna of different water bodies have revealed a decline in fish diversity,96 especially those of threatened species.¹¹⁵ Therefore for conservation of the river system allochthonous sources of pollution like sewage, dumping of garbage, mining and agricultural activities needs to be monitored. Different conservation measures like using a species as flagship species, creating awareness and starting different projects towards conservation needs to be adopted,¹¹⁶ also small hydropower projects should be prioritised over large reservoir-based hydropower projects since they are more environmentally friendly and have fewer negative effects on flora and wildlife.117

Research Gap and Future Prespective

Only a small number of the area's waterbodies have been thoroughly examined; the remainder of the wetlands and many lentic water bodies have mostly remained uncharted due to accessibility concerns (remote location), financial limitations, and the locals' intense religious convictions. Fish production in the area can increase significantly when the existing resources (waterbodies and fish fauna) are used wisely in order to meet the demands of UT's growing population. As the majority of the water bodies in the Jammu and Kashmir region are unexplored, concealing a substantial portion of the fish flora and its gene pool, an integrative strategy can assist close the study gap.

Therefore a collaboration on the molecular aspect of fisheries in J and k especially in Jammu region with conventional taxonomy will immensely help in better understanding of the fish ecology of the region and will also aid in properly identifying and conserving the gene pool thereby boosting the growth of the economically important fishes belonging to this region.

Acknowledgments

The authors acknowledge the support and guidance received from the Department of Zoology, Central University of Jammu, J&K. Anchal Chib (NET-JRF) is also thankful to University Grants Commission for providing Junior Research Fellowship.

Funding

The author(s) received no financial support for the research, authorship, and/or publication of this article.

Conflict of Interest

The authors declare that there is no conflict of interest.

References

- Paller VGV, Corpuz MNC, Ocampo PP (2013). Diversity and Distribution of Freshwater Fish Assemblages in Tayabas River, Quezon (Philippines). *Philippine Journal of Science* 142 (1): 55-67.
- Cadrin SX, Silva VM (2005). Morphometric variation of yellowtail flounder. *ICES Journal of Marine Science*, 62(4), 683–694. doi:10.1016/j.icesjms.2005.02.006.
- Chaklader R, Bakar Siddik MA, Nahar A (2015) Taxonomic diversity of paradise threadfin polynemus paradiseus (Linnaeus, 1758) inhabiting southern coastal rivers in Bangladesh. Sains Malaysiana, 44(9), 1241–1248. doi:10.17576/jsm2015-4409-04.
- Siddik MAB, Hanif MA, Chaklader MR, Nahar A, Mahmud S (2015) Fishery biology

of gangetic whiting Sillaginopsis panijus (Hamilton, 1822) endemic to Ganges delta, Bangladesh. *Egyptian Journal of Aquatic Research*, 41(4), 307–313. doi:10.1016/j. ejar.2015.11.001.

- Tripathy SK(2020)Significance of Traditional and Advanced Morphometry to Fishery Science. Journal of Human, Earth, and Future Vol. 1, No. 3, September, 2020 http:// dx.doi.org/10. 28991/HEF-2020-01-03-05.
- Strauss RE, Bond CE (1990) Taxonomic methods: morphology. In: Schreck CB, Moyle PB (eds) Methods for fish biology. American Fisheries Society, Maryland, pp 109–140
- Uiblein F (1995) Morphological variability between populations of Neobythites stefanovi (Pisces:Ophidiidae) from the deep Red

Sea and the Gulf of Aden. Marine Ecology Progress Series, 124, 23–29. doi:10.3354/ meps124023.

- Sarder MRI, Simonsen V (2006). Final report on the study and workshop on causes for and consequences of reduced growth in Indian major carps (p. 62).
- Hurlbut T, ClayD (1998). Morphometric and meristic differences between shallowand deep-water populations of white hake (Urophycis tenuis) in the southern Gulf of St. Lawrence. Canadian Journal of Fisheries and Aquatic Sciences, 55(10), 2274–2282. doi:10.1139/f98-110.
- Turan C (2004).Stock identification of Mediterranean horse mackerel (Trachurus mediterraneus) using morphometric and meristic characters. ICES *Journal of Marine Science*, 61(5), 774–781. doi:10.1016/j. icesjms.2004.05.001.
- Teletchea F(2009) Molecular identification methods of fish species: reassessment and possible applications. Rev Fish Biol Fisheries (2009) 19:265–293 DOI 10.1007/s11160-009-9107-4.
- Allendorf FW, Phelps SR (1988) Loss of genetic variation in hatchery stock of cutthroat trout. Trans. Am. Fish. Soc. 109: 537-543.
- Wimberger PH(1992) Plasticity of fish body shape - the effects of diet, development, family and age in two species of Geophagus (Pisces: Cichlidae). *Biol. J. Linn. Soc.* 45: 197-218.
- Swain DP, Ridell BE, Murray CB (1991) Morphological differences between hatchery and wild populations of coho salmon (Oncorhynchus kisutch): environmental versus genetic origin. Can. J. Fish. Aquat. Sci. 48: 1783-1791.
- Hossain MAR, Nahiduzzaman M, Saha D, Habiba Khanam MU, Alam MS (2010). Landmark-based morphometric and meristic variations of the endangered carp, kalibaus Labeo calbasu, from stocks of two isolated rivers, the Jamuna and Halda, and a hatchery. Zoological Studies, 49(4), 556–563.
- Juliao (2020)https://agencia.fapesp.br/ environmental-factors-influence-fishmorphology-and-behavior/32635/.
- 17. Radojkovic N, Marinovic Z, Miloskovic A, Radenkovic M, Duretanovis S, Lujic J,

Simic V (2018) Effects of Stream Damming on Morphological Variability of Fish: Case Study on Large Spot Barbell Barbus balcanicus.

- Eklöv P, Svanbäck R (2006). Predation risk influences adaptive morphological variation in fish populations. In American Naturalist (Vol. 167, Issue 3). American Nationals. doi:10.1086/499544.
- Keskin E, Atar HH. DNA barcoding commercially important fish species of Turkey. Molecular ecology resources. 2013 Sep;13(5):788-97.
- Cohen NJ, Deeds JR, Wong ES, Hanner RH, Yancy HF, White KD, Thompson TM, Wahl M, Pham TD, Guichard FM, Huh I, Austin C, Dizikes G, Gerber SI (2009) Public health response to pufer fsh (tetrodotoxin) poisoning from mislabeled product. *J Food Prot* 72:810–817
- Bashir A, Bisht SB, Mir IJ, Patiyal SR, Kumar R (2016) Morphometric variation and molecular characterization of snow trout species from Kashmir valley, India. Mitochondrial DNA part A DNA Mapping, Sequencing, and Analysis Volume 27, 2016 – Issue 6.
- Straüss RE, Bond CE. In: Methods for Fish Biology, 1990. Amer Fish Soc. Schreck CB, Moyle PB, editors. Maryland, U.S.A: Bethesda; 1990. Taxonomic methods: morphology; pp. 125–130.
- 23. Ko HL, Wang TY, Chiu TS, Lee MA, Leu MY, Chang KZ, ChenWY, Shao KT (2013) Evaluating the Accuracy of Morphological Identification of Larval Fishes by Applying DNA Barcoding, PLoS ONE 8(1): e53451. https:// doi.org/10.1371/journal.pone.0053451.
- 24. Zhang JB, Hanner R. DNA barcoding is a useful tool for the identification of marine fishes from Japan. Biochemical Systematics and Ecology. 2011 Feb 1;39(1):31-42.
- 25. Uiblein F. Morphological variability between populations of Neobythites stefanovi (Pisces: Ophidiidae) from the deep Red Sea and the Gulf of Aden, Marine ecology progress series. vol 124:23-29,1995.
- Tillett BJ, Field IC, Bradshaw CJA, Johnson G, Buckworth RC, Meekan M G, Ovenden JR(2012). Accuracy of species identification by fisheries observers in a north Australian

shark fishery. *Fisheries Research*, 127-128, 109-115. https://doi.org/10.1016/j. fishres.2012.04.007

- CohenJN, DeedsRJ, WongSE, HannerHR, YancyFH, WhiteDK, ThompsonMT, Wahll, PhamDT, GuichardMF, Huhl, Chandra S, Barat A, Singh M, Singh K B, Matura R (2012) DNA Bar-Coding of Indian Coldwater Fishes of Genus Schizothorax (Family: Cyprinidae) from Western HimalayaWorld Journal of Fish and Marine Sciences 4 (4): 430-435, 2012
- Austin C, Dizikes G, Gerber IS (2009) Public Health Response to Puffer Fish (Tetrodotoxin) Poisoning from Mislabeled Product *J Food Prot* (2009) 72 (4): 810–817.
- 29. Ghouri MZ, Ismail M, Javed MA, Khan SH, Munawar N, Umar AB, Nisa M, Aftab SO, Amin S, Khan Z and Ahmad A (2020) Identification of Edible Fish Species of Pakistan Through DNA Barcoding. Front. Mar. Sci. 7:554183. doi: 10.3389/fmars.2020.554183
- Cawthorn MD, Steinman AH, Witthuhn CR (2012) DNA barcoding reveals a high incidence of fish species misrepresentation and substitution on the South African market. Food research international Volume 46, Issue 1Pages 1-436.
- Vartak RV, Rajendran N, Lakra SW(2014) DNA barcoding: Importance in *fisheries* research and food safety, *BioTechnology An Indian Journal* volume 9.
- Callejas C, Ochando MD (2001) Molecular identification (RAPD) of the eight species of the genus Barbus (Cyprinidae) in the Iberian Peninsula. *J Fish* Biol 59:1589–1599. doi:10.1111/ j.1095-8649.2001.tb00223.x.
- Gewin V (2002) All living things, online. Nature 418: 362-363.
- Wilson EO (2003) What is Nature Worth For? Part 1: Span ZLIV-4: 54-57 Part 2: XLIV, 23-29.
- Simonetti JA (1997) Biodiversity and a taxonomy of Chilean taxonomists. Biodiversity and Conservation 6: 633-637.
- Tahseen Q (2014) Taxonomy-The Crucial yet Misunderstood and Disregarded Tool for Studying Biodiversity. J Biodivers Endanger Species 2: 128. doi:10.4172/2332-2543.1000128
- 37. Wheeler QD, Raven PH ,Wilson EO (2004) Taxonomy: Impediment or expedient?

Science, 303 (5656), 285.

- De Carvalho MR, Bockmann FA, Amorim DS, de Vivo M, et al.(2005). Revisiting the taxonomic impediment. Science, 307, 353.
- Talwar PK, Jhingren AG(1991) Inland Fishes of India and adjacent Countries. Oxford and IBH Publishing Co. Pvt. Ltd., New Delhi. Vol. II:115-116.
- 40. Jayaram, K C(2010). Freshwater fishes of the Indian region. Narendra Pub. House.
- 41. Sarma K J, Mankodi P C (2017). Deciphering identification of inland fishes of Gujarat using dna barcoding. *Turkish Journal of Fisheries and Aquatic Sciences*. 17. 1059-1061. 10.4194/1303-2712-v17_5_22.
- Hebert PD, Cywinska A, Ball SL, deWaard JR. Biological identifications through DNA barcodes. *Proc Biol Sci.* 2003 Feb 7;270(1512):313-21. doi: 10.1098/ rspb.2002.2218. PMID: 12614582; PMCID: PMC1691236.
- Brasier MJ, Wiklund H, NealL, Jeffreys R, Linse K, Ruhl H, Glover AG (2016)DNA barcoding uncovers cryptic diversity in50% of deep-sea Antarctic polychaetes.R.Soc. opensci.3: 160432.
- 44. Korshunova T, Picton B, Furfaro G et al.(2019) Multilevel fine-scale diversity challenges the 'cryptic species' concept. Sci Rep 9, 6732 (2019). https://doi.org/10.1038/s41598-019-42297-5
- 45. The Global Taxonomy Initiative .2020: A Stepby-Step Guide for DNA Barcoding
- Kochzius M, Seidel C, Antoniou A, Botla SK, Campo D, Cariani A, et al. (2010) Identifying Fishes through DNA Barcodes and Microarrays. PLoS ONE 5(9): e12620.
- Fischer J (2013) Fish identification tools for biodiversity and fisheries assessments: review and guidance for decision-makers. FAO Fisheries and Aquaculture Technical Paper No. 585. Rome, FAO. 2013. 107 pp.
- Gregory TR. DNA barcoding does not compete with taxonomy. Nature. 2005 Apr 28;434(7037):1067. doi: 10.1038/4341067b. PMID: 15858548.
- Ward RD, Zemlak TS, Innes BH, Last PR, Hebert PDN. DNA barcoding Australia's fish species. Philosophical transactions of the Royal Society of London. Series B. 2005; 360(1462):1847-1857. https://doi.

org/10.1098/rstb.2005.1716.

- 50. Moura T, Silva MC, Figueiredo I, Neves A, Munoz PD, Coelho MM and Gordo LS (2008) Molecular barcoding of north-east Atlantic deep-water sharks: species identification and application to fisheries management and conservation Marine and Freshwater Research 59(3) 214-223
- Hubert N, Hanner R, Holm E, Mandrak NE, Taylor E, et al. (2008) Identifying Canadian Freshwater Fishes through DNA Barcodes. PLOS ONE 3(6): e2490. https:// doi.org/10.1371/journal.pone.0002490
- Holmes, Bronwyn, Steinke, Dirk, Ward, Robert. (2009) Identification of shark and ray fins using DNA barcoding. *Fisheries Research*. 95. 280-288. 10.1016/j.fishres.2008.09.036.
- 53. Steinke D, Zemlak TS, Boutillier JA, Hebert PND (2009)DNA barcoding of pacific canadas fishes, Marine Biology. November 2009 156:2641–2647
- Ahmed MS, Dina SR, Nahar L, Islam NN, Reza HA (2018) Molecular characterization of Channa species from Bangladesh based on Cytochrome C Oxidase Subunit I (COI) gene FishTaxa (2018) 3(4): 87-93.
- 55. Lakra SW, Verma MS, Goswami M, Lal KK, Mohindra V, Punia P, Gopalakrishnan AK, Singh KV, Ward RD, Hebert P (2011) DNA barcoding Indian marine fishes. Molecular Ecology Resources (2011) 11, 60–71
- Chandra S, BaratA, Singh M, Singh KB, Matura R (2012) DNA Bar-Coding of Indian Coldwater Fishes of Genus Schizothorax (Family: Cyprinidae) from Western HimalayaWorld *Journal of Fish and Marine Sciences* 4 (4): 430-435, 2012
- 57. Barman D, Roy S, Kumar V, Mandal SC, Kumar V (2012) DNA barcoding applications in aquaculture, Global Aquaculture Advocate Saturday, 3 March 2012.
- Khedkar GD, Jamdade R, Naik S, David L, Haymer D (2014). DNA barcodes for the Flshes of the Narmada, one of India's longest rivers. PLoS ONE., 9(7)
- Thapliyal M, Pokhriyal H, K Sati B, Nagpure N, Singh M, and Thapliyal A (2015) Molecular characterization of coldwater fishes of district Uttarkashi, Uttarakhand using DNA Barcoding Environment Conservation Journal 16(3) 109-116.

- Kumar S, Singh D.Genetic and morphometric comparison of two isolated populations of Barilius bendelisis (Cypriniformes: Cyprinidae) from the Indian Himalayas. Revista de Biología Tropical vol. 67, no. 3, 2019, pp. 466-477
- Kundu S, Chandra K, Tyagi K, Pakrashi A, Kumar V (2019) DNA barcoding of freshwater fishes from Brahmaputra River in Eastern Himalaya biodiversity hotspot . Mitochondrial DNA part B 2019, vol. 4, no. 2, 2411-2419.
- Alam A, Chadha KN, Kumar PA, Chakraborty KS, Joshi DK, Sawant BP, Das SCS, Kumar J, KumarT (2019) DNA Barcoding and Biometric Investigation on the Invasive Oreochromis niloticus (Linnaeus, 1758) from the River Yamuna of Uttar Pradesh.Indian *Journal of Animal Research*.2020.(54):856-863.
- Kashmiri Pandit network: Geography of Jammu and Kashmir State, http://ikashmir. net/geography/chapter1.1.html
- 64. Nasim and keng (2012) Directory of lakes and waterbodies of j&k state using remote sensing & GIS technology. department of environment and remote sensing sda colony bemina srinagar / paryawaran bhawan, forest complex, jammu.
- Heckel JJ (1838) Fische aus Caschmir. Karl Freiherrn von Hugel. Wien. Gedruckt Bei Den P. P. Mechitaristen. 112pp+13pl
- 66. Day F (1876) On the fishes of Yarkand. Proc Zoology Soc London:781–807
- 67. Silas EG (1960) Fishes from Kashmir Valley. J Bombay Nat Hist Soc 57(1):145–182
- Das SM, Subla BA (1964) The Ichthyofauna of Kashmir, Part II. The speciation of Kashmir fishes. Ichthyologica 3:57–62.
- Das SM, Nath S (1965) The ichthyofauna of Poonch Valley (J&K). Kashmir Sci 2(1-2):149–155
- Yousuf AR (1996) Fishery resource of Kashmir. In: Khan AH, Pandit AK (eds) Ecology, Environment and Energy. Kashmir University, Srinagar, pp 75–120
- Kullander SO, Fang F, Delling B, Ahlander E (1999) The fishes of the Kashmir Valley. pp. 99-162. In: Lennart Nyman (Ed.), River Jhelum, Kashmir Valley, impact on the aquatic environment SWEDMAR, Göteborg.
- 72. Enderlin O, Yousuf AR (1999) The

environmental impacts f the Uri hydropower project on the fish community in the river Jhelum. In: Nyman L (ed) River Jhelum, Kashmir valley: impacts on the aquatic environment. SWEDMAR, Göteborg, pp 169–186

- Balkhi MH (2007) Fish diversity of Jammu & Kashmir and its conservation. In: Patloo (ed) Kashmir speaks. G. M. Publishers, Kashmir, pp 104–118
- 74. Khan I, Ali M (2013)Current Status of the Fish Fauna of River Jhelum, Kashmir, J&K, Open Access Scientific Reports 2: 694
- 75. Jan R , Gupta R , Najar MA and Zuber SM (2015) Abiotic Status of River Jhelum with Special Reference to its Ichthyofaunal Diversity Research Journal of Animal, Veterinary and Fishery Sciences. ISSN 2320 – 6535
- Ahmed I, Ahmad Z, Ahmad I (2017) Current status of fish fauna of river Jhelum and dal lake of kashmir valley Bulletin of Pure and Applied Sciences .Vol 36 A (Zoology), No.2, P.85-92.
- Bahir M, Chauhan R, Mir FM, Ashraf M, Amin N(2016) Effect of pollution on fish diversity in Anchar lake Kashmir,International journal of fisheries and aquatic resources 2017; 5(1): 105-107
- Brraich SO, Malik AI (2016) Fish fauna of Wular Wetland (Ramsar site in Kashmir Himalayas, India) Iran. J. Ichthyol. (September 2016), 3(3): 218–221.
- Rumysa K, Sharique AA, Bilal A, Tariq Z and Farooq M (2016) On the fish diversity, conservational management and rehabilitation aspects of Wular Lake, Kashmir India. Ecological Communication.*Biosci. Biotech. Res.* Comm. 9(4): 872-877.
- Qadri S, Bhat FA, Balkhi MH, Shah TH, Hussain N, Bhat AB, Altaf M, and Aalia S (2018) Fish biodiversity, catch composition, distribution pattern of fishes in Wular Lake and their conservational measures for sustainable fish production. *Journal of Pharmacognosy and Phytochemistry* 2018; 7(6): 1883-1887.
- Mushtaq AS, Balkhi MH, Abubakr A, Kumar A, Shah HT, Ahmed B, Shah F, Talia S, QadriS, Farooq I (2020) Current status of fish fauna, catch composition of Hokersar Wetland, Kashmir, International Journal of

Fauna and Biological Studies 2020; 7(1): 15-18

- Hamid J, Singh RK (2019) Diversity of Fishes in River Viashaw: the left tributary of river Jhelum of Jammu and Kashmir, India, JETIR may 2019, volume 6, issue 5.
- Rashid G,Singh R.(2020)Present status of fish diversity and their conservational measures for sustainable fishproduction in the vaishav stream of kashmir himalaya, india, Journal of critical reviews,vol 7, issue 16, 2020
- Akhtar T, Ali G (2016) DNA barcoding of Schizothorax species from the Neelum and Jhelum Rivers of Azad Jammu and Kashmir Mitochondrial DNA Part B • December 2016.
- 85. Bashir A, Bisht BS, Mir JI, Kumar R, Patiyal RS. Morphological, molecular characterization and taxonomic status of *Triplophysa marmorata* and Triplophysa kashmirensis (Cypriniformes: Nemacheilidae) from Kashmir valley, India. Rev Biol Trop.2016 Jun;64(2):473-82. doi: 10.15517/ rbt.v64i2.19591. PMID: 29451748.
- Das SM and Nath S (1966) The icthyofauna of Jammu province (J&K), *Kashmir Sci.*,2 (1-2) : 65-78.
- Das SM and Nath S (1971) A revision of the fishes from Jammu province, Kashmir Sci., 7: 1-12.
- Malhotra YR, Jyoti MK, Dutta SPS (1975) An aid to the identification of fishes found in Jammu division of J&K State. Jammu Univer Rev 5:50–66
- 89. Joshi CB, Sehgal K L, Sunder S (1981) Observations on the fishery resources of the hill streams of Jammu Province with special reference to mahseer and other commercially important species. Indian journal of fisheries
- 90. Tilak 1971. The fishes of river tawi and its r-tributaries (jammu & kashmir state) with notes on ecology.ZoologicalSurvey of India, Calcutta 183-217.
- Dutta SPS and Malhotra YR (1984) An upto date checklist and a key to identification of fishes of Jammu. Jammu Univ. Review., 2 : 65-92.
- Jyoti MK, Sharma K, Gupta K, Baba DI (2006). Aquatic biodiversity: a review of freshwater flora and fauna of J&K State. In: Jyoti MK, Sharma KK, Gupta K (eds)

Proceeding National Symposium: status of coldwater fishes with reference to fragile Himalayan aquatic ecosystems. Classic Printers, Bari Brahmna, pp 258–292

- Baba DI, Sharma KK, & Jasrotia S (2014) Ichthyofaunal Diversity of River Chenab: A Case Study. *International Journal Of Lakes and Rivers*, 55-62.
- 94. Bhutyal R, Langer S (2015) Current status of fish fauna of river Chenab, in kishtwar district, j&k, Asian academic research *journal of multi disciplinary*, Issue 5.
- 95. Gandotra R, Zaman U R, Vivek (2017). Longitudinal Patterns of Population Structure for Fishes Inhabiting River Tawi in Jammu region (J&K),Biosci.,Biotech. Res. Asia,Vol.14(3), 1121-1127. http://dx.doi. org/10.13005/bbra/2550
- 96. Sharma A, Dutt SPS (2012) Present and past status of fish fauna of river Basantar, an important tributary of the river Ravi, in Samba district, Jammu (J&K). *Journal of Applied and Natural Science* 4 (1): 123-126
- Vohra A, Jyoti MK, Gupta K (2013)Some ornamental fishes of Jammu region International journal of innovative research and development. Vol 2 Issue 8: 379-390.
- Arif M, Gandotra R, Choudhary N, Sharma D, Lone A, Dogra A (2019) Study on Distribution and Abundance of Indigenous Ornamental Fishes from Water Bodies of Jammu (J&K), Biosciences Biotechnology Research Asia. Vol. 16(2), p. 483-489
- 99. Dutta SPS (2015) Survey and systematic analysis of Fish fauna of erstwhile Udhampur District, Jammu region, with new records of Barilius radiolatus Gunther, Botia lohachata Chaudhari, Botia dario (Ham. Buch.) and Glyptothorax punjabensis Mirza and Kashmiri, Environment Conservation Journal 16(3) 39-47, 2015.
- Rathore V, Dutta SPS (2015) Fish fauna of river Ujh, an important tributary of the river Ravi, District Kathua, Jammu, Environment Conservation Journal 16 (1&2)81-86.
- 101. Khajuria B, Langer S, Khan S (2015) Icthyodiversity and Species Richness of Chadwal Stream, Kathua District (Jammu & Kashmir), International journal of scientific research. Volume - 4 | Issue : 10,63-65.
- 102. Gandotra R, Sharma P (2015) Study of

Ichthyofaunal Diversity in a stream in Sunderbani, District Rajouri, Jammu (J&K), International Journal of Multidisciplinary Research and Development Volume: 2, Issue: 9, 401-404.

- 103. Kour H , Sharma KK, Sharma A (2015) Icthyofaunistic survey of some lotic water bodies of R.S Pura, Tehsil, Jammu, Journal of International Academic research for Multidisciplinary, 3: 2320-5083.
- 104. Dutta SPS (2016) Fish fauna of Wajoo nullah, an important tributary of the river Ravi in Kathua District, Jammu Region, Jammu and Kashmir State, India. Journal of Applied and Natural Science 8 (2): 1087 - 1089
- 105. Hussain S, Dutta SPS (2016)Fish fauna of tehsil mendhar, district- poonch, jammu region, j&k state, with a new record of botia birdi, ompok pabda and glyptothorax punjabiensis from poonch district, Proc.Zool. Soc.India. 15 (1): 1 – 11
- 106. Khajuria B, Langer S, Kumar S, Choudhary A(2016).Fish diversity and habitat relationship of selum nullah and Aik nullah with reference to physico chemical parameters at Bari Brahmana,Jammu district,JandK.Journal of advanced scientific research,2016,7(1):53-59
- 107. Nisa RU, Gupta K, Wani SM, Allie KA, kouser N (2020) Study on diversity and ecology of ichthyofauna of Rajouri district, Jammu and Kashmir, India.Rec. zool. Surv. India: Vol. 120(4):363–372
- 108. Dutta SPS (2021) Fish Fauna of the River Ravi and Its Some Tributaries with a New Record of Ailia puncata and Clupisoma naziri for Punjab State and Union Territory of Jammu and Kashmir, India. Aquaculture and Fisheries Studies.vol3,issue1:1-5.
- 109. Gupta SC and Dutta SPS (2021) Fish Fauna of River Sewa, an Important Himalayan Tributary of the River Ravi, in Kathua District of Union Territory of Jammu & Kashmir, India, biomedical journal of scientific and technical research. Volume 36- Issue 3:28474-28478.
- 110. Arif M and Gandotra R (2017) DNA barcoding of ornamental fishes of Jammu:7th international barcode of life conference 2017 nov.
- Kadye WT, Moyo NAG, Magadza CHD, Kativu S (2008)Stream fish assemblages in relation to environmental factors on

a Montane Plateau(Nyika Plateau,Malawi). Environmental biology of fishes,83,417-423.

- 112. Makori AJ, Abuom PO, Kapiyo R,et al.(2017) Effects of water physico-chemical parameters on tilapia (Oreochromis niloticus) growth in earthen ponds in Teso North Sub-County, Busia County. Fish Aquatic Sci 20, 30. https:// doi.org/10.1186/s41240-017-0075-7
- 113. Krishan R (2016)Impact of climate change on fish fauna of jammu and Kashmir,International journal of innovative research in science and engineering.vol no 2,Issue 03,March .
- 114. Bhat FA, Yousuf AR, Balkhi MH (2020). Diversity of Fishes in Jammu and Kashmir State. In: Dar, G., Khuroo, A. (eds) Biodiversity

of the Himalaya: Jammu and Kashmir State. Topics in Biodiversity and Conservation, vol 18. Springer, Singapore. https://doi. org/10.1007/978-981-32-9174-4_33

- Andotra P (2014) Impact of pollution on water quality and fishes of river Tawi. Ph.D. Thesis, University of Jammu, Jammu 2014.
- 116. Bhatt JP, Pandit KM (2015) Endangered Golden mahseer Tor putitora Hamilton: a review of natural history, Rev Fish Biol Fisheries 26:25–38
- 117. Sharma KA, Thakur NS (2017) Assessing the impact of small hydropower projects in Jammu and Kashmir: A study from northwestern Himalayan region of India.