

Assessment of Phenolic Compounds in the Surface Waters of Godavari Canal, Andhra Pradesh, India

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ABSTRACT

The present study was intended to determine the concentrations of phenolic compounds in surface waters of Godavari canal by molecular spectrophotometry. Samples were collected from fifteen sampling stations using grab sampling method for a period of four months (from November to February) at 10 day intervals. Total phenols in water samples were determined using molecular spectrophotometry after distillation, complexation with 4-aminoantipyrine and extraction into chloroform. The concentration of phenolic compounds was reported in the range of 80-179 mg/lit, well within the limits considering the earlier reports in the literature and the standards prescribed by Central Pollution Control Board (CPCB) of India. Relatively higher levels of phenolic compounds (100-179 mg/lit) were reported in nine out of fifteen sampling locations. Statistical analysis showed that there is a significant difference in the concentrations of phenolic compounds in the month of November with respect to January (at $p = 0.05$) and November with respect to February (at $p < 0.05$).

Key words: Phenolic compounds, Surface waters, Spectrophotometry, Distillation.

INTRODUCTION

Phenolic compounds are ubiquitous in environment. Many of them are found in nature and found to be responsible for colour of many flowers and fruits¹. They are undegradable organic materials and pollute the natural ecosystem. They can arise from natural substance degradation, industrial activity and agricultural practices. They are found in wastes of synthetic resin, plastics, rubber-proofing, dye manufacturing and many chemicals². They are found to be toxic and persistent. They are proved to have bioaccumulation effects in animal and vegetable organisms and may be dangerous for human health³.

There are several anthropogenic sources for phenolic compounds in the environment.

The presence of phenolic compounds in aquatic environment may be due to human and animal metabolism, industrial activities and agricultural practices⁴. Polyphenolic macro molecules are also present in some species of marine algae, including green macro algae and red macro algae⁵. Low levels of phenol are found in some foods (smoked summer sausage, fried chicken, some species of fish, cheese) and in tobacco smoke.

Phenolic compounds in water environment may have a natural, industrial, domestic or agricultural origin. In agricultural practice, phenolic compounds are employed as herbicides, insecticides or can derive from degradation of the chlorophenoxy-carboxylic herbicides and organophosphorous insecticides⁶. Phenolic compounds can reach the water environment through industrial

and domestic waste and through treated sewage discharges. Once in water reservoir, phenols may undergo destruction and transformation by the impact of different physico-chemical factors and by the activity of aqueous organisms like fungi, algae, saprophytes⁷. The presence of phenols in a stream is undesired because of their strong action, persistence and their toxicity to invertebrates⁸.

A number of analytical methods are reported for the determination of phenols, based on spectrophotometric, fluorimetric, kinetic, gas chromatographic, liquid chromatographic and enzyme and biosensors. Due to several characteristic features such as high sensitivity, operational facility and low cost the UV-visible spectroscopy is one of the most useful analytical techniques⁹.

In the recent years, many papers have been devoted to the occurrence of phenols and phenolic compounds in surface waters of canals¹⁰⁻¹¹. However, there is paucity of data about their concentration levels in surface waters of canals in India. Hence present study was carried out to investigate the concentrations of phenolic compounds in Godavari canal. In the present study surface water samples were collected for a period of four months period at ten day intervals and the levels of phenolic compounds (in mg/lit) were determined using molecular spectrophotometry method prescribed by American Public Health Association (APHA)¹².

MATERIALS AND METHODS

Fifteen sampling stations were chosen based on human activity to collect the surface water samples from the canal as shown in Figure 1.

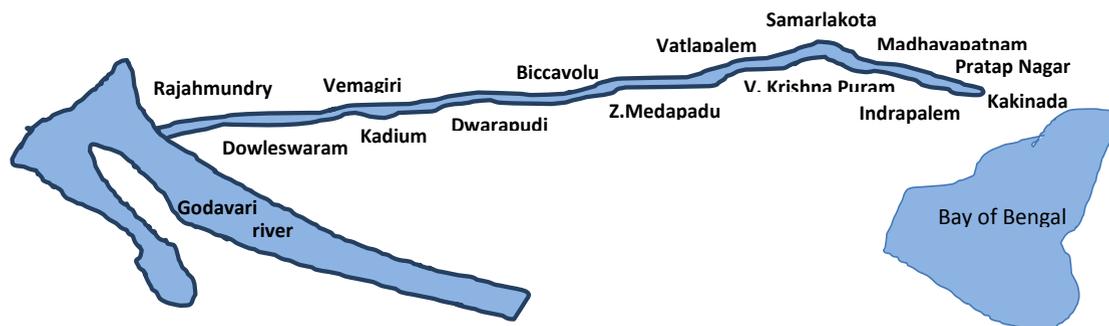


Fig. 1: Location map showing the sampling stations

The geographical location of the sampling sites is presented in Table 1.

Grab sampling method was employed to collect the samples randomly from all the fifteen stations. The sampling was carried out over a period of four months at ten day intervals to give a total number of 180 samples. The samples were collected and conserved as reported in the literature. The samples were collected in 1 litre amber glass bottles and acidified with 4 ml of concentrated sulphuric acid and were kept cool during transportation and then stored at 4°C, until analysed. The samples were stored not more than 24 hours in the refrigerator. Most of the samples were analysed within a few hours after they were brought to the lab.

The analytical procedure recommended by APHA (American Public Health Association) was used to determine the concentration of phenolic compounds¹². The levels of phenolic compounds (in mg/lit) were determined using molecular spectrophotometry after distillation, complexation with 4-aminoantipyrine and extraction into chloroform. The phenolic compounds react with 4-aminoantipyrine in the presence of potassium ferricyanide at a pH of 10 to form a stable reddish-brown antipyrine dye. The amount of colour produced is a function of the concentration of phenolic compounds. JASCO V-450 spectrophotometer was used to carry out the spectrophotometric studies.

RESULTS AND DISCUSSION

The mean concentrations of phenols in four different months are presented in Table 2

The variation in the concentration of phenolic compounds in the sampling stations during the four month period is shown in Figure 2. The levels of phenolic compounds were found to be in the range of 80-179 mg/lit in the sampling stations.

Higher levels of phenolic compounds (in the range of 110-179 mg/lit) were reported in

seven sampling stations, sampling stations 1 to 4, sampling station 13, sampling station 14 and sampling station-15. The higher levels of phenolic compounds in these stations may be attributed to more urbanized activities and agricultural activities near these stations. In these sampling stations the density of population was found to be high.

Table 1: The geographical location of the sampling sites

S.No	Name of the sampling station	GPS location of sampling station	
SS1	Rajahmundry	17°00'03"N	81°79'83"E
SS2	Dowleswaram	16°95'63"N	81°78'97"E
SS3	Vemagiri	16°93'79"N	81°80'73"E
SS4	Kadium	16°90'87"N	81°82'27"E
SS5	Dwarapudi	16°92'08"N	81°91'44"E
SS6	Biccavolu	16°96'95"N	82°05'04"E
SS7	Z.Medapadu	17°00'46"N	82°10'22"E
SS8	V. Krishna Puram	17°01'44"N	82°19'21"E
SS9	Vatlapalem	17°02'56"N	82°13'07"E
SS10	Samarlakota	17°05'28"N	82°17'92"E
SS11	Madhavapatnam	16°99'34"N	82°20'27"E
SS12	Sarpavaram	16°99'40"N	82°21'06"E
SS13	Pratap Nagar	16°97'48"N	82°22'05"E
SS14	Indrapalem	16°96'71"N	82°21'60"E
SS15	Kakinada	16°94'64"N	82°23'36"E

Table 2: The mean concentrations of phenols in four different months

S.No	Sampling station	Phenol Concentration in mg/L			
		November	December	January	February
SS1	Rajahmundry	115	118	122	125
SS2	Dowleswaram	113	116	118	113
SS3	Vemagiri	107	112	112	116
SS4	Kadium	105	107	112	116
SS5	Dwarapudi	74	77	97	87
SS6	Biccavolu	91	95	102	101
SS7	Medapadu	81	78	82	87
SS8	Vetlapalem	94	98	92	105
SS9	Samarlakota	89	94	103	99
SS10	V. Krishna Puram	83	88	94	97
SS11	Sarpavaram	93	98	99	107
SS12	Madhavapatnam	91	95	98	99
SS13	Pratap Nagar	110	114	117	117
SS14	Indrapalem	109	113	113	111
SS15	Kakinada	114	118	119	123

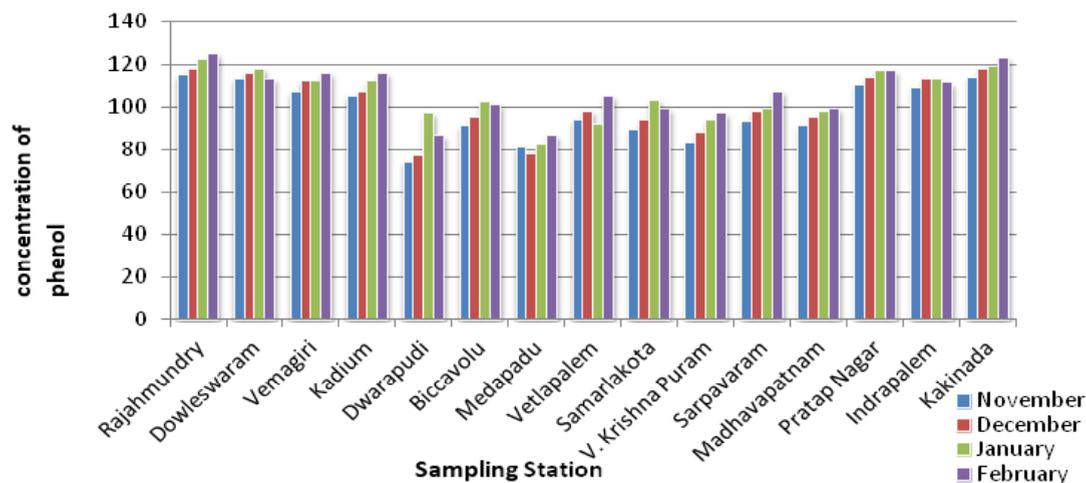


Fig. 2: The variation in the concentration of phenolic compounds in the sampling stations

The phenol concentrations in all the sampling locations were well within the permissible limits prescribed by the Central Pollution Control Board (CPCB) of India. Statistical analysis showed that there is a strong correlation between concentrations of phenolic compounds determined in the months of November and December ($r = 0.99$), November and January ($r = 0.90$), November and February ($r = 0.95$). A strong correlation was also observed in the months of December and January ($r = 0.91$), December and February ($r = 0.96$) and January and February ($r = 0.88$). T-test was carried out to determine the significant difference in the concentrations of phenolic compounds in different months. There is a significant difference in the concentrations of phenolic compounds in the month of November with respect to January (at $p = 0.05$) and November with respect to February (at $p < 0.05$).

CONCLUSION

The APHA recommended molecular spectrophotometric method for determination of

phenolic compounds in surface waters of Godavari canal was used. The study shows that the surface waters of Godavari canal were polluted with phenolic compounds mainly due to rapid urbanisation and anthropogenic activities. Even though the levels of phenolic compounds were lower than permissible limits prescribed by Central Pollution Control Board of India, continuous monitoring is required due to the fact that continuous emissions of phenolic compounds in surface waters results in their deposition and their increase in soil sediment.

Due to the lack of previous data on the levels of phenolic compounds in surface waters of Godavari canal, the results of the present study could not be compared and correlated to determine the extent of contamination of the surface waters of the canal in the past. However the data in the present study may serve as a reference in the future since regular monitoring of phenolic compounds is essential due to their toxicity and bioaccumulation effects in animal and vegetable organisms.

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